



Food and Agriculture  
Organization of the  
United Nations



ورشة عمل خطط الطوارئ كتمرين محاكاة عملي

Contingency Exercise Workshop

*Xylella fastidiosa*

Hammamet, Tunisia, 26 – 28 May 2025

# Aphrophoridae candidate vectors of *Xylella fastidiosa* in Tunisia

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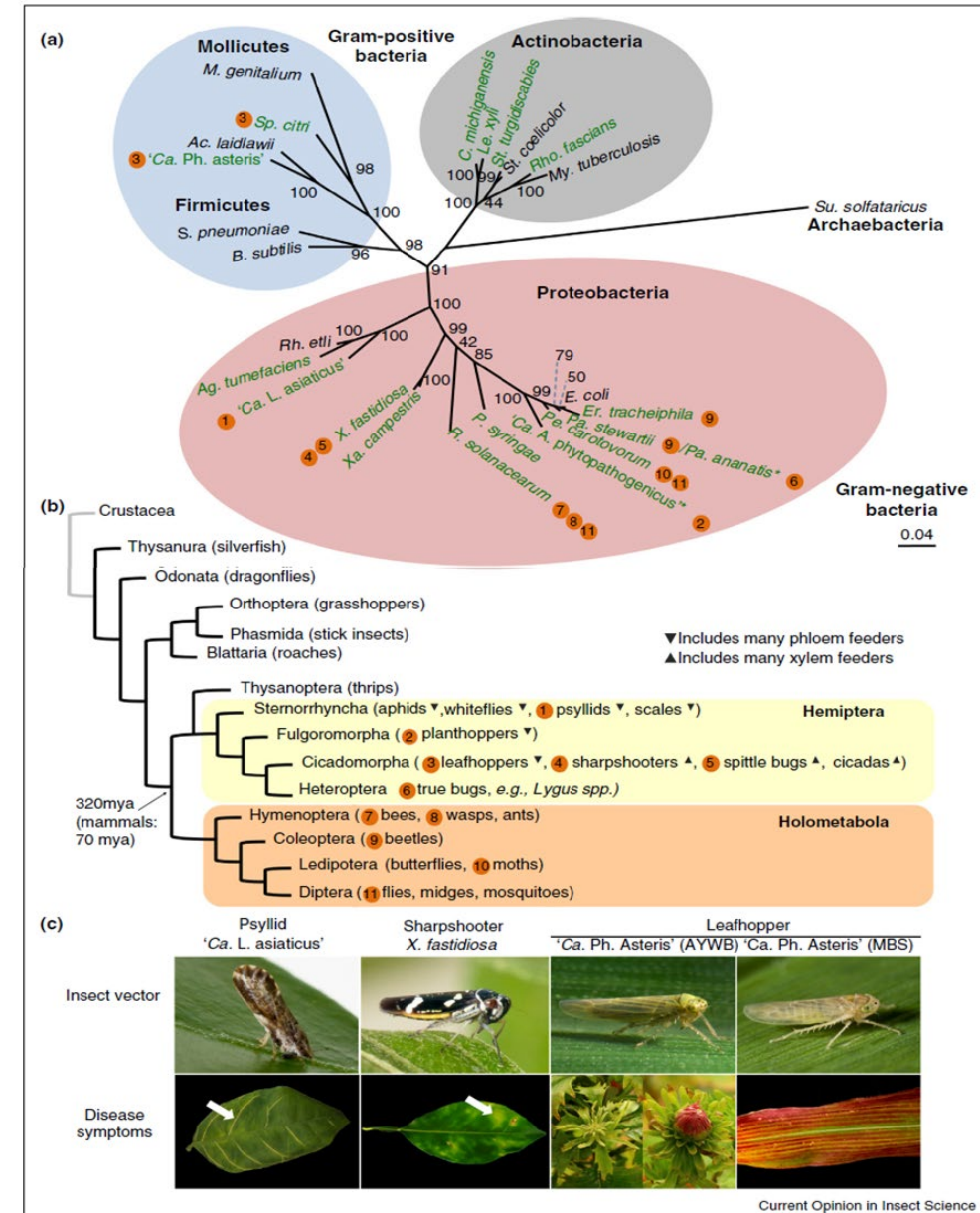


# Vectors/pathogens

Several bacteria are transmitted by insect vectors (psyllids, thrips, leafhoppers, spittlebugs, butterflies, beetles, etc.)

## *Xylella fastidiosa*

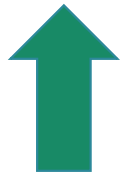
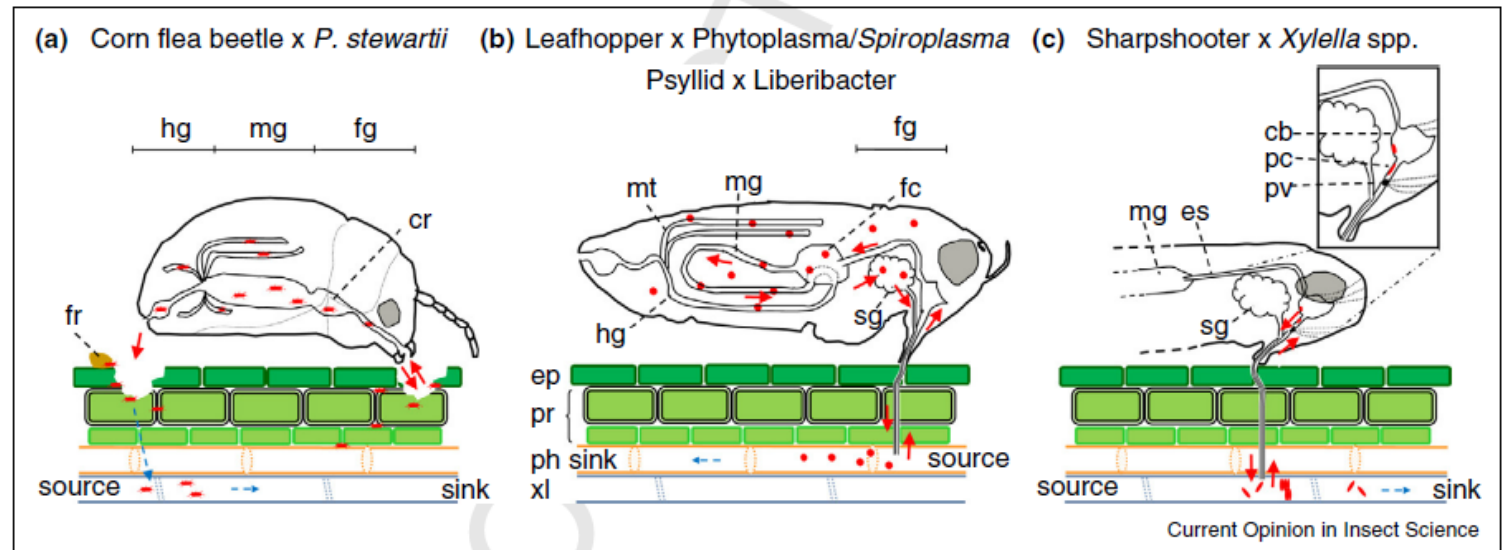
- Gram-negative bacterium
- Transmitted by spittlebugs and leafhoppers



### 3 strategies of bacterial transmission by insects

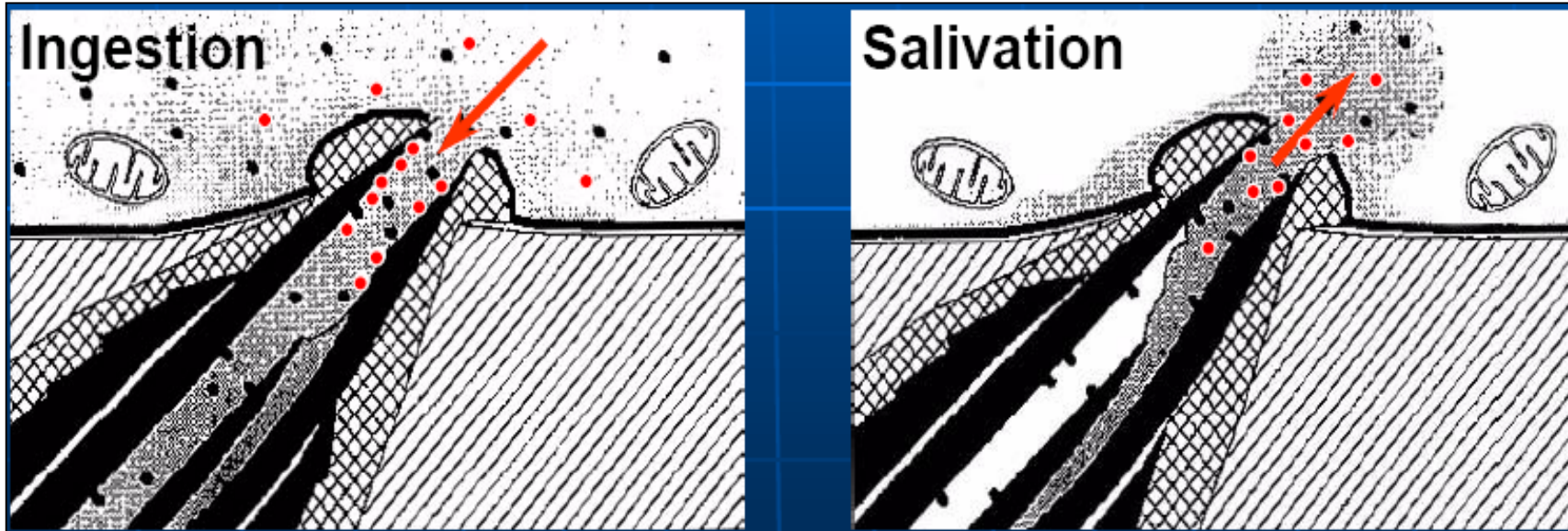
Xylem, phloem, parenchyma feeders

- *X. fastidiosa* non-circulating but persistent in the vector
- Bacteria attached to the cuticle in the cibarium, multiplies and is inoculated by egestion





# *Xylella fastidiosa* acquisition/transmission



The adult, by feeding on the sap of an infected plant, becomes immediately capable of transmitting the disease (Rodas, 1994).

As the bacterium multiplies in the insect's oral cavity, the insect remains infectious for several months (Purcell and Finlay, 1979; Hill and Purcell, 1995).

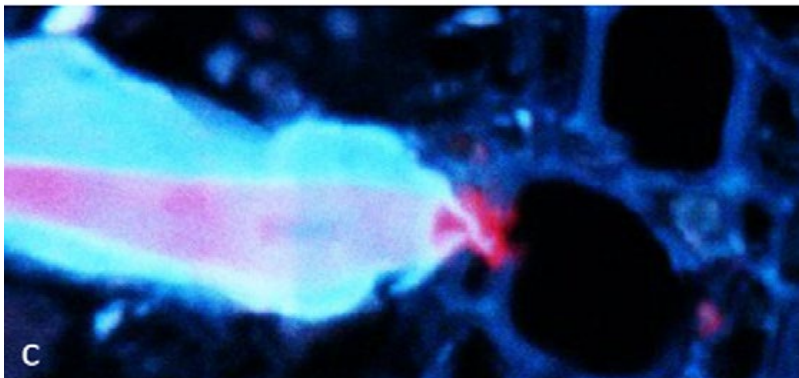
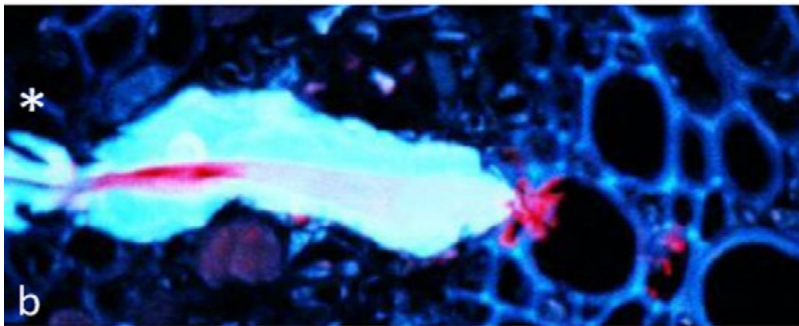
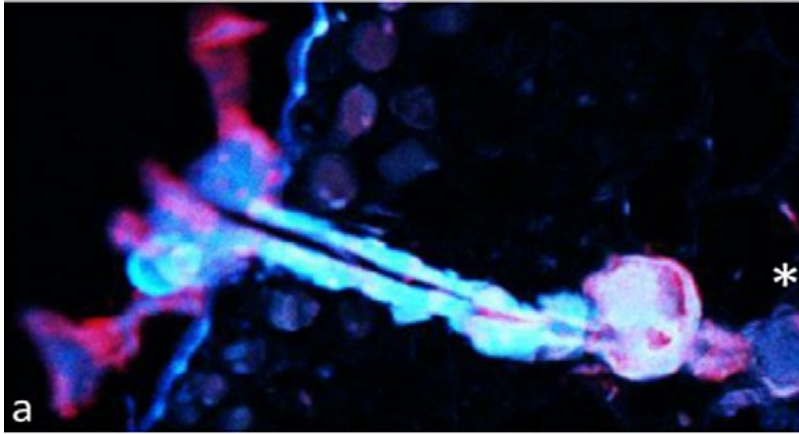
**The presence of a few bacterial cells is enough for the insect to transmit the disease.**



## *X. fastidiosa*

- The threshold value for bacterial acquisition by the insect is around  $10^6$  to  $10^7$  bacterial cells per gram of tissue (Hill and Purcell, 1997).
- Acquisition efficiency is positively correlated with the feeding duration on the host plant (Almeida and Purcell, 2003; Daugherty and Almeida, 2009).
- Transmission rates of *X. fastidiosa* is positively correlated with the number of vectors (Costa et al., 2000).

# *Homalodisca vitripennis* (Germar)/ *Xylella fastidiosa*



A xylem-limited bacterium that causes Pierce's disease (grapevine)

inoculation of *X. fastidiosa* during vector feeding

Scanning microscopy showing a salivary sheath of *H. vitripennis* on a vine petiole. (a) Distal half of the salivary sheath on the surface of the petiole. (b) and (c) adjacent sections showing the proximal half of the sheath within the petiole as its tip enters a xylem vessel.



# Auchenorrhyncha potential vectors in the world

Currently, the group includes **70** Hemiptera species

- Aphrophoridae
- Cercopidae
- Clastopteridae
- Cicadellidae
- Cicada
- Membracidae

# Auchenorrhyncha potential vectors in the world

- 70 species in Americas
- 5 in Asia
- 11 in Europe
- 7 in Mediterranean region

North  
Africa

- *Philaenus tessellatus*\*
- *Philaenus maghresignus*
- *Neophilaenus campestris*\*
- *Neophilaenus linneatus*
- *Lepeyronia coleoptrata*
- *Aphrophora alni*
- *Graphocephala hirsuta*



# Main Vectors feeding on xylem

17 suspected species

- **Cicadellidae** : *Cicadella viridis*, *Graphocephala virsuta*, *Homalodisca vitripennis*, *Oncometopia nigricans*
- **Clastopteridae** : *Clastoptera achatina*
- **Aphrophoridae** : *Aphrophora alni*, *Aphrophora salicina*, *Lepyronia coleoptrata*, *Neophilaenus campestris*, *N. lineatus*, *Philaenus spumarius*, *P. italosignus*, *P. tessellatus*, *P. maghresignus*, *Cercopis vulnerata*, *C. sanguinolenta*

**Confirmed vectors in mediterranean region (Italy):**  
*Philaenus spumarius*, *Neophilaenus campestris*, *P. italosignus*

# Auchenorrhyncha vectors and potential vectors in the world



*Homalodisca  
vitripennis*



*Lepyronia  
quadrangularis*



*Clastoptera  
achatina*



*Oncometopia  
nigricans*



*Graphocephala  
virsuta*



*Aphrophora  
alni*



*Aphrophora  
salicina*



*Philaenus  
spumarius*



*Neophilaenus  
lineolatus*



*Neophilanus  
campestris*



*Cercopis  
sanguinolenta*



*Cercopis  
vulnerata*



*Cicadella  
viridis*



*Philaenus  
tessellatus*



*Philaenus  
italosignus*

More than 17 species are vectors or potential vectors

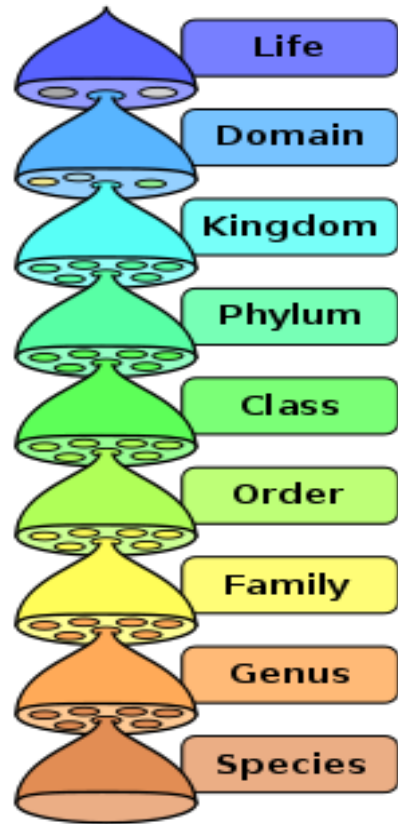
*Philaenus spumarius* is a well-known vector, extremely polyphagous species (nearly 1000 plant species) and can be found in large numbers in many environments (hedges, meadows, crops, gardens, river banks, high-altitude meadows, forests).

In America, the most important vectors of *X. fastidiosa* are *G. virsuta*, *O. nigricans*, *P. spumarius* and *H. vitripennis*

***Graphocephala virsuta* reported from Algeria**



# Taxonomy of the most efficient vector



Life

Domain

Kingdom

Phylum

Class

Order

Family

Genus

Species

Animalia

Arthropoda

Insecta

Hemiptera/Auchenorrhyncha

Aphrophoridae

**Philaenus**

***Philaenus spumarius***

**Neophilaenus**

***Neophilaenus campestris***



(Spittlebugs 5,3 à 6,9 mm)

# Spittlebugs and Leafhoppers Collection Methods

- Sweep net
- Threshing with a Japanese umbrella
- Mechanical vacuum cleaners (D-Vac)
- Sticky yellow chromo-attractant panels (glue traps)
- Traps embedded in the ground (Barber traps).
- Light traps, Malaise tent type traps are also available
- Sticky Wire Traps

Each technique produces different results regarding the species captured and the combination of several techniques is necessary to exhaustively identify and evaluate Auchenorrhyncha populations.

# Trap types



Malaise tent



Sweep net



D-Vac



Malaise trap



Japanese umbrella



Sticky yellow traps



Light traps



Piège à fils gluants  
Cliché J.-L. Dommanget

Sticky Trap

# Important Notations

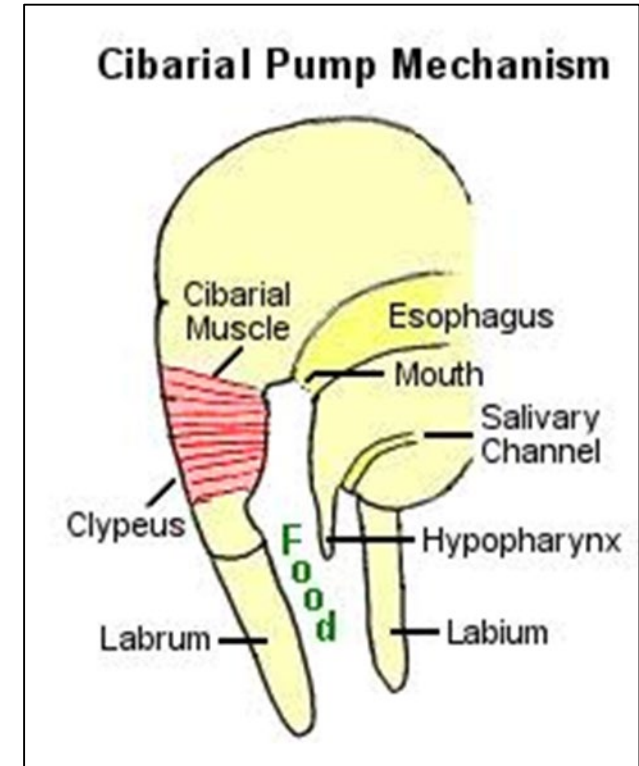
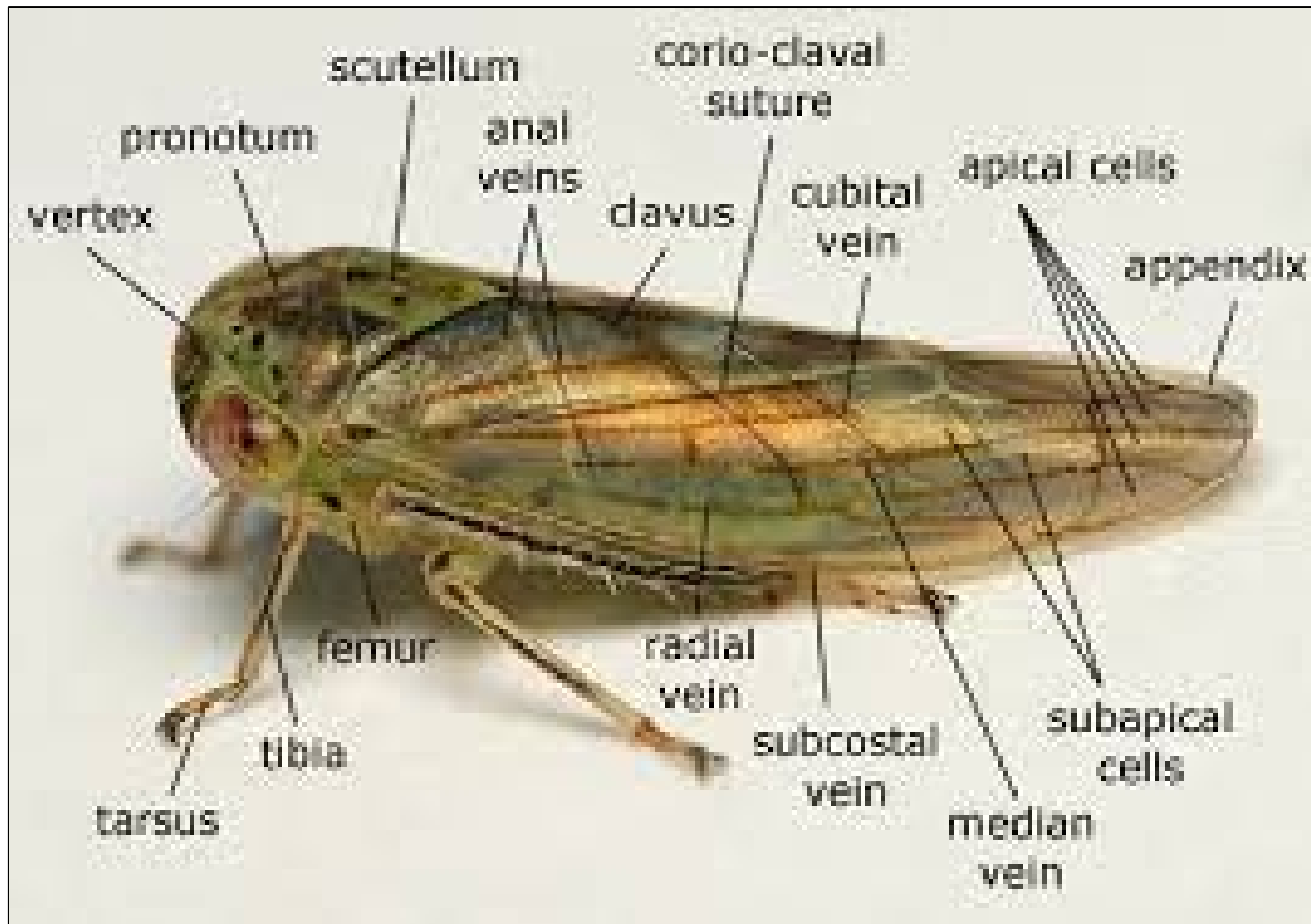
Collection monitoring data must include:

- 1) A unique collection number;
- 2) geographic coordinates;
- 3) The date and name of the person who collected samples;
- 4) The name of the host plant;
- 5) A list of plants; a typology of plant density showing symptoms. ;  
collection of insect vectors by sweep net.

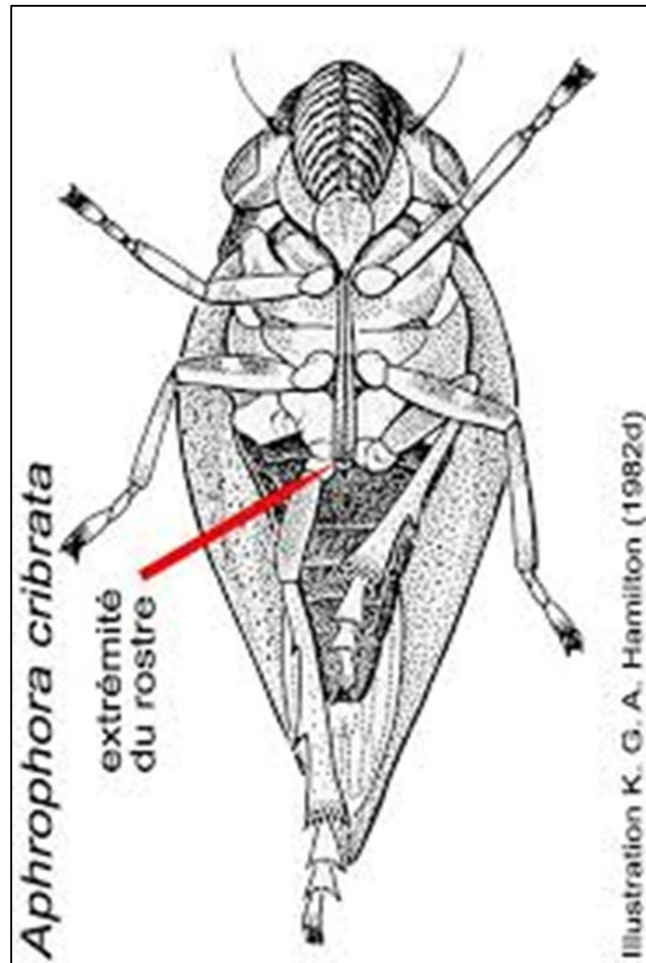




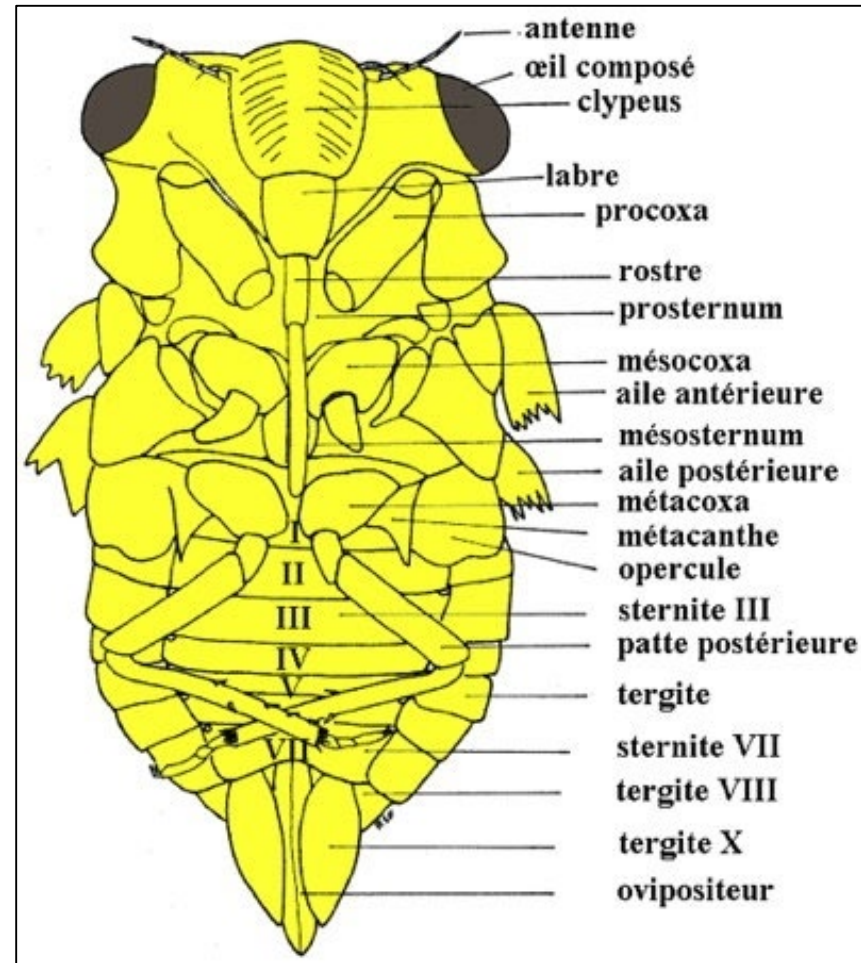
# Identification criteria (dorsal view)



# Morphological features (ventral view)



Male

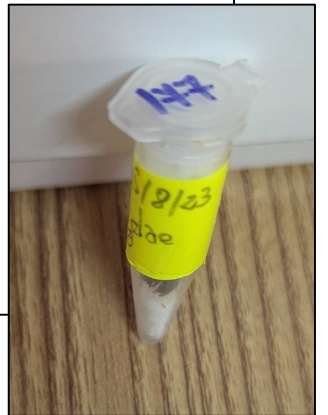


Female

Male genitalia used for species identification

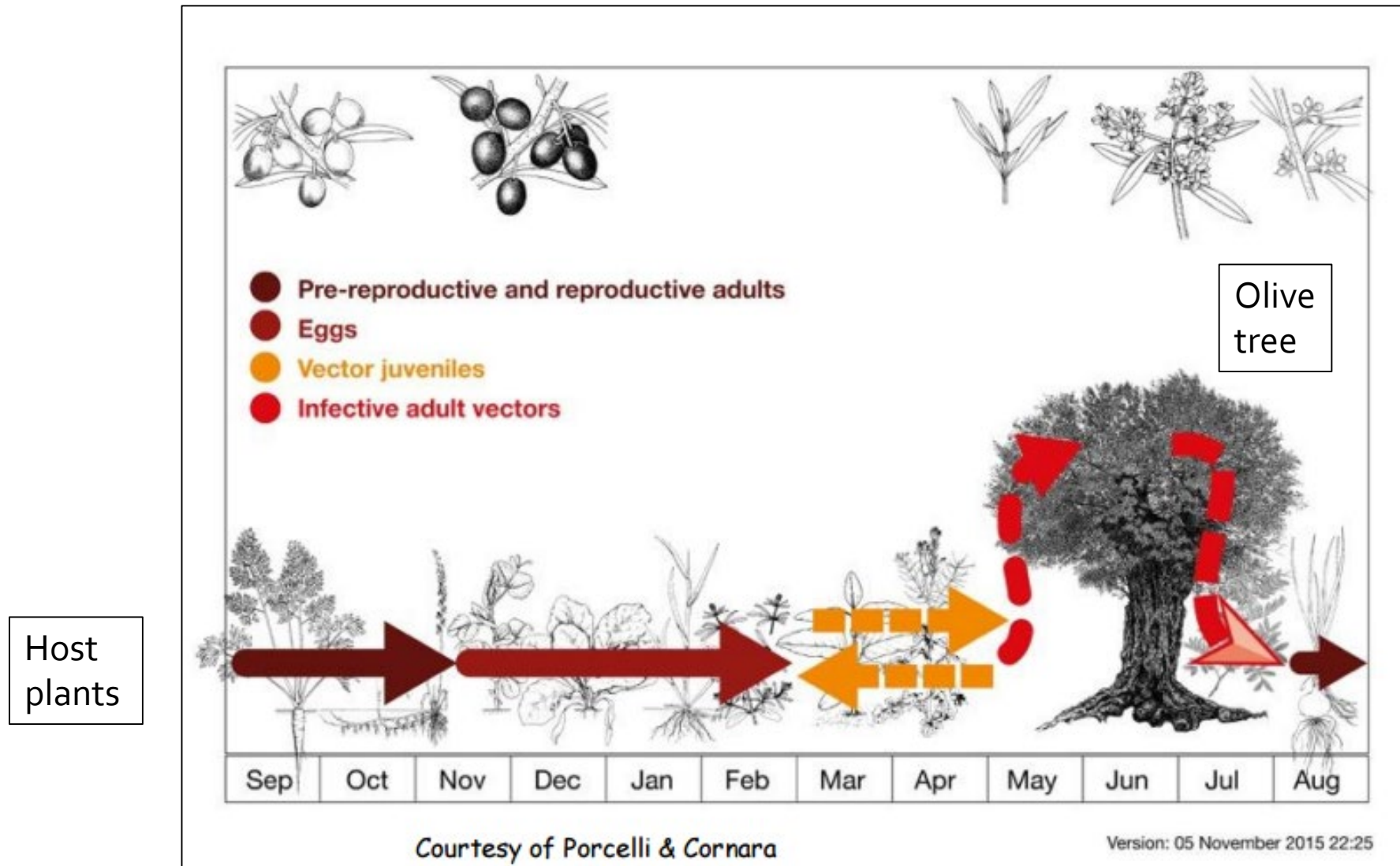
# Insect Conservation

- The captured insects must be placed in ethanol 75 then transferred to ethanol 96 in containers which prevent evaporation of the solvent.
- The volume of insects should not exceed one third of the volume of ethanol.
- The storage can be done at 4°C for a short period or at -20°C for a longer period
- The tubes must be legibly and precisely labeled.





# Philaenus cycle and Xylella dissemination



# Tunisian experience

- Collection and Identification of potential vectors
- Forest/Dry grassland/Olive/Citrus/Peach/Almond propections
- Host plants of vectors
- Bar-coding and Xylella detection
- Natural ennemies identification



# Spittlebugs collecting regions in Tunisia with the focus on two reservoir forests



Feija forest



Dar Chichou forest

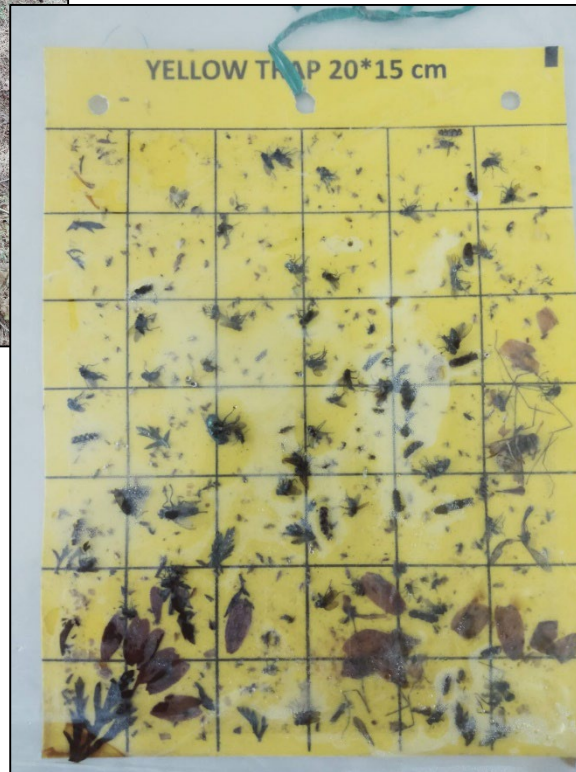


## Collecting spittlebugs

- Sweep-net (50 sweeps in forests, 20 under fruit trees)
- Plant (20 plants of each species)



# Trap used



- Limited numbers
- Insects adhered strongly to the glue, making removal difficult



- Good method













# Identified material

More than 8,000 Auchenorrhyncha captured by sweep net

- 5010 Aphrophoridae (4 species identified)
- 3082 Cicadellidae (129), Typhlocibidae (5), Caliscelidae (1), Issidae (9), Delphacidae (3), Cixiidae (2), Dictyopharidae (2), Flattidae (1), Tettigometridae (6), Membracidae (2)



## 4 Aphrophoridae : *Philaenus* and *Neophilaenus* identification features

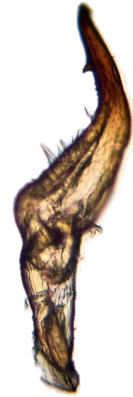
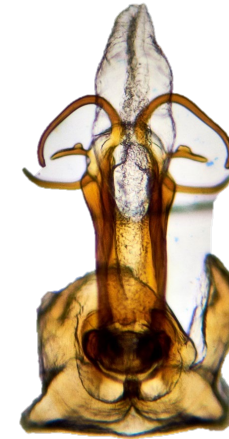
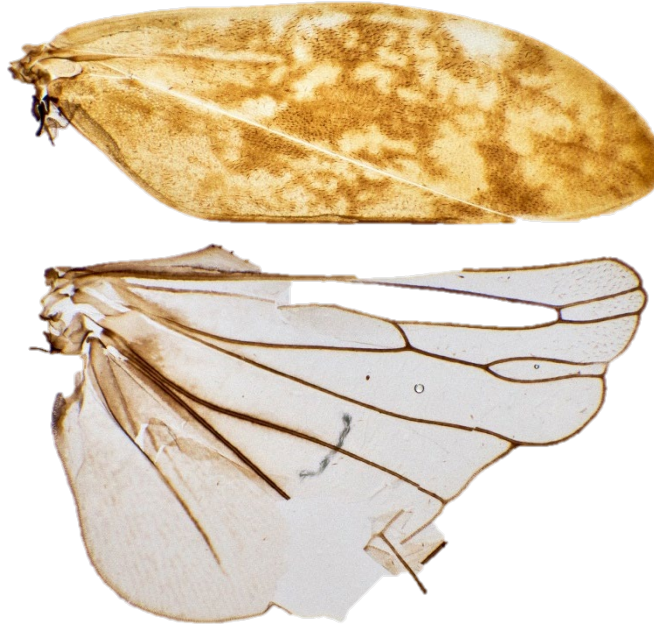
Species	Adult	Aedeagus	Hind leg
<i>P. maghresignus</i>			
<i>P. tessellatus</i>			
<i>N. campestris</i>			
<i>N. lineatus</i>			

We did not identify  
*Philaenus spumarius*  
in Tunisia



# Insect identification : *Philaenus tessellatus*

- Morphological: insects captured as males dissected to observe their genitalia

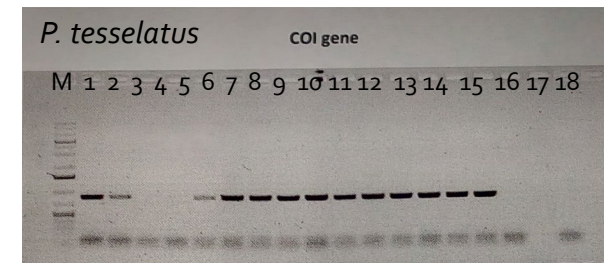


- Molecular: COI-based bar-coding

>16\_COI-

```
FTTTGGGTATAATTTATGCTATAATAGCAATTGGTTTATTAGGTTTTGTGGTTTGAGCTCATCATATTTTACTGTAGGTAT  
AGATGTTGATACACGTGCATATTTTACTTCAGCCACAATAATTATTGCTGTACCTACGGGTATCAAAATTTTATGTTGATT  
GGCTACAATACATGGAATACCATTCAAATTGNCTTCTCCTATTTTATGATCAATTGGATTGTATTTNT
```

Sequence very close to GU446992.1 from NY Museum with a bootstrap value of 99,13% (NCBI bank)



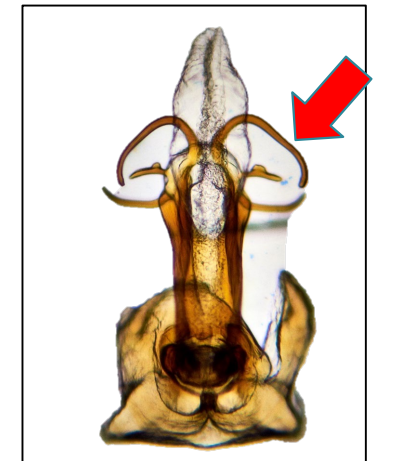
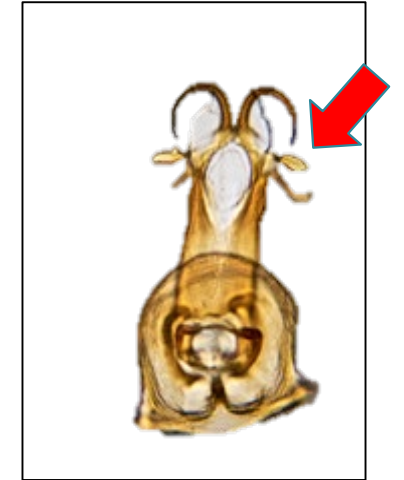


# Discrimination with *Philaenus spumarius*



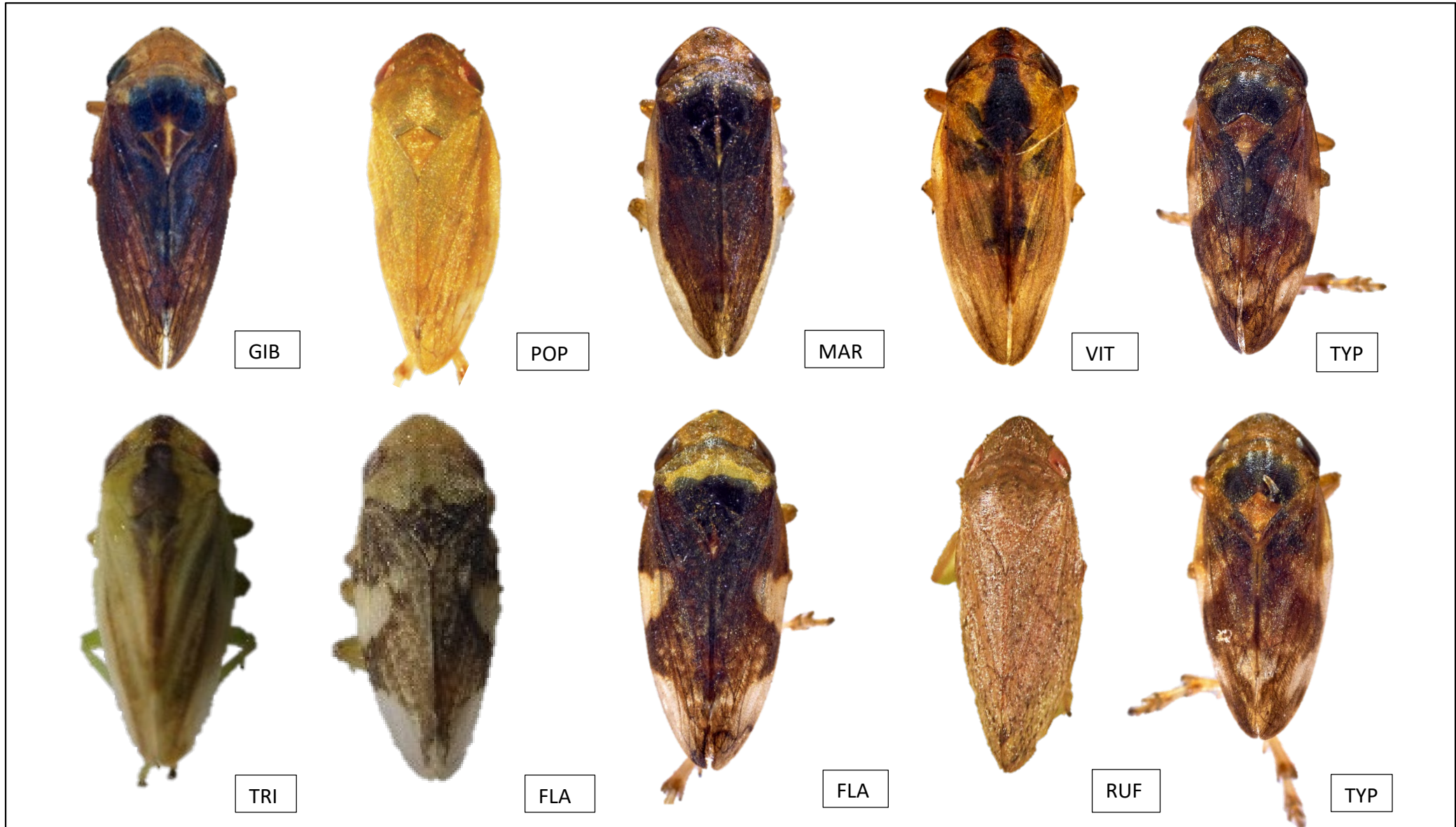
***Philaenus Spumarius*** (Linnaeus), A, Male posterior and anterior view; B, Hind tibia spines; C, forewing; D, hind wing; E, Male genital capsule lateral view; G, right style ventral-dorsal aspect; J, aedeagus, dorsal view;

***Philaenus Spumarius***



***Philaenus tessellatus***

# Morph diversity of *Philaenus tessellatus*



Morphs TRI, GIB, MAR, VIT, TYP, POP, FLA, RUF and TRI



# Intermediate morphs



MAR  
/FLA



VAR/  
TYP



VIT/  
TRI?



TRI ?

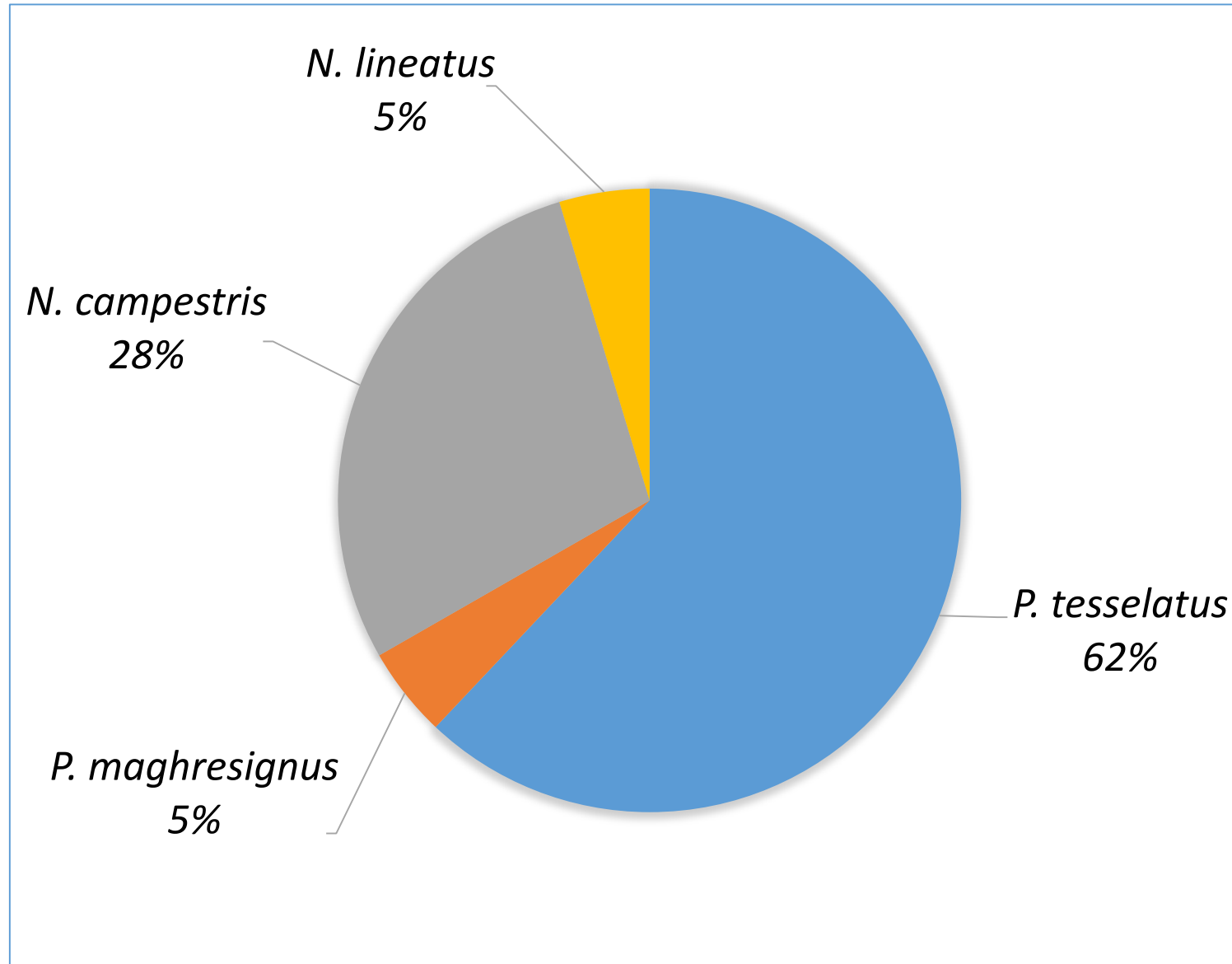


VIT ?



VIT ?

# Composition of Aphrophoridae adults in tunisian regions, 2018-2021

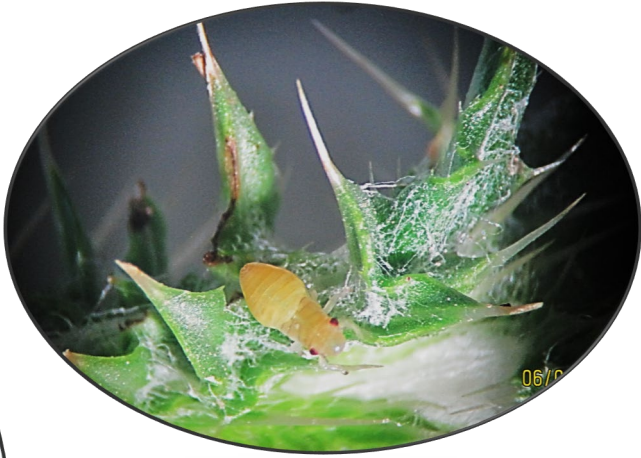




# Biological cycle of *P. tessellatus*



Fev.-Mar.



Nymphs



molting  
Nymphs



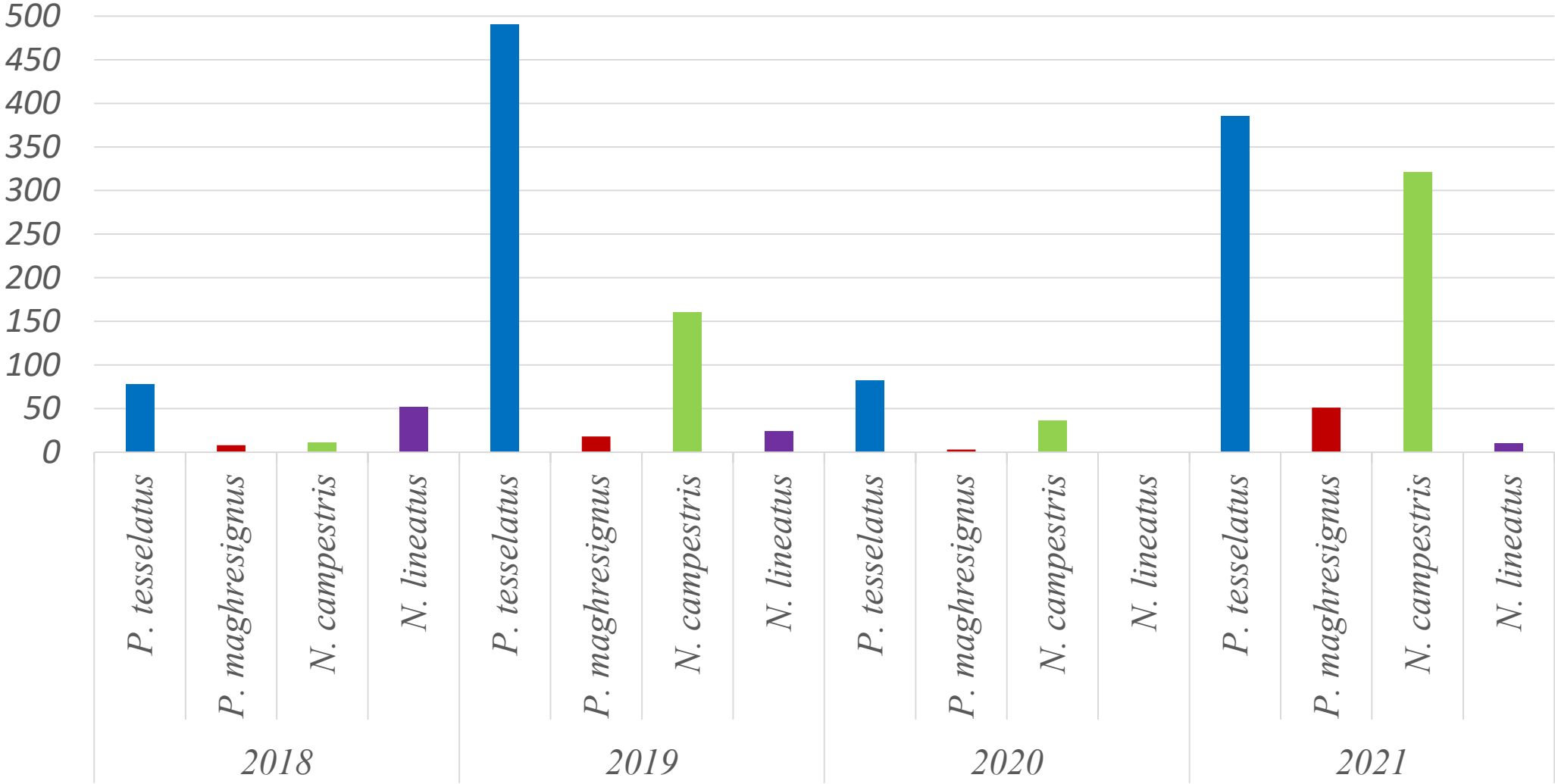
Adults

Late Apr.-May

Mar.-Apr.



# Frequency variations of the four spittlebugs according to years



# *Philaenus tessellatus*/*Neophilaenus campestris*/*P. maghresignus*



Polyphagous and well adapted at many plants and habitats

- *Sonchus oleraceus*
- *Smyrnum olusatrum*
- *Glebionis segetum*
- *Cirsium arvense*
- *Ranunculus repens*
- *Sonchus oleraceus*
- *Scolymus grandiflorus*

- *Rumex crispus*
- *Daucus carota*
- *Mercurialis annua*
- *Hedysarum coronarium...*

Oligophagous on Poaceae

- *Festuca*
- *Avena sativa*
- *Bromus...*

Monophage

*Asphodelus microcarpus*

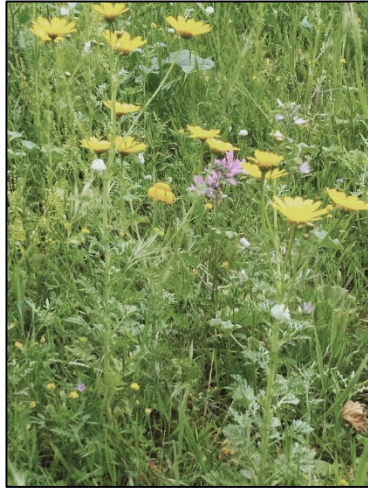


# Host plants/Nymphs-Adult of *P. tessellatus* (Dar Chichou)

*Scolymus grandiflorus*



*Glebionis segetum*



*Sonchus oleraceus*



*Rubia perigrina*



*Daucus carota*



*Cirsium arvense*

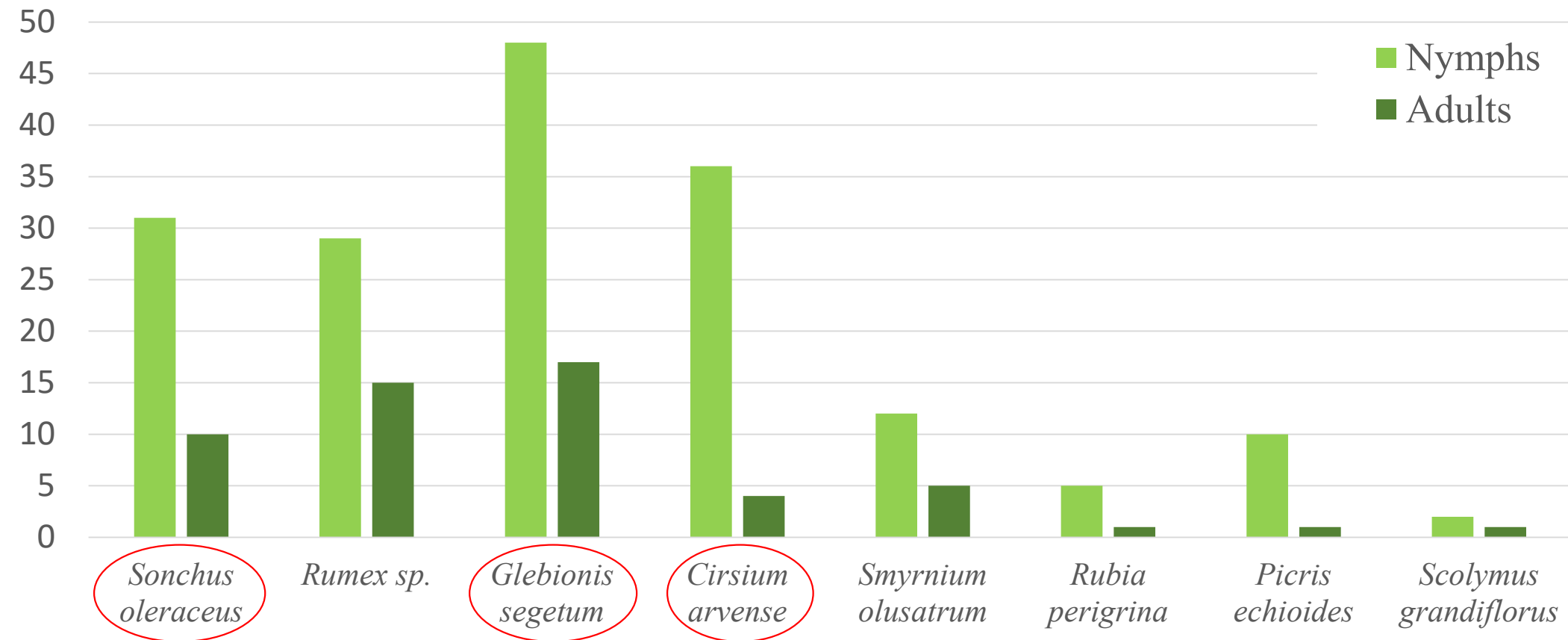


*Smyrniurn olusatrum*





Spittlebug nymphs and adults of *Philaenus tessellatus* on diverse weeds, Dar Chichou April 2021



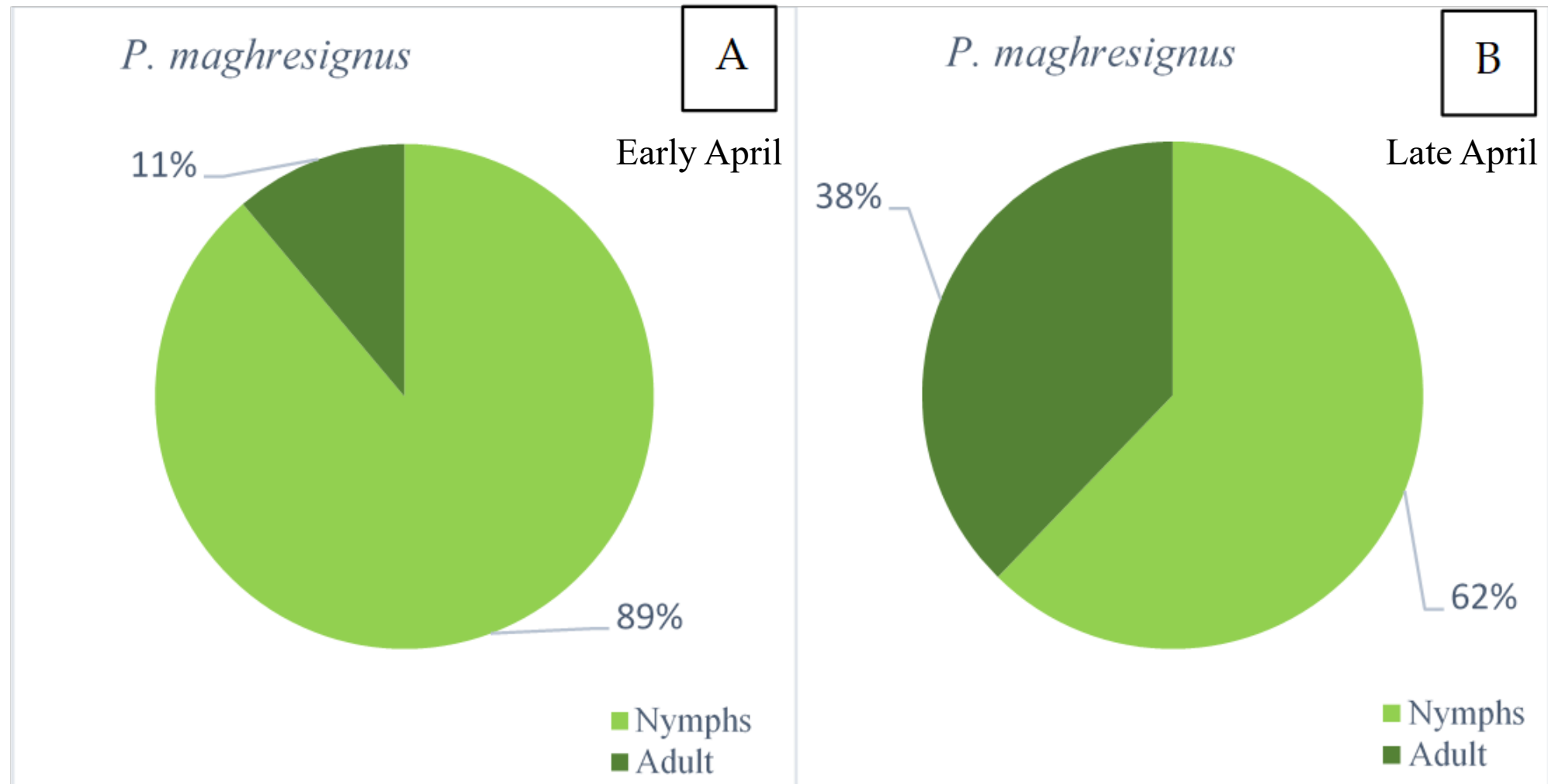
# Nymphs of *P. maghresignus* on *Asphodelus* plant collected from Feija



Monophagous



# *P. maghresignus* nymphs and adults on *Asphodelus microcarpus*, Feija 2021

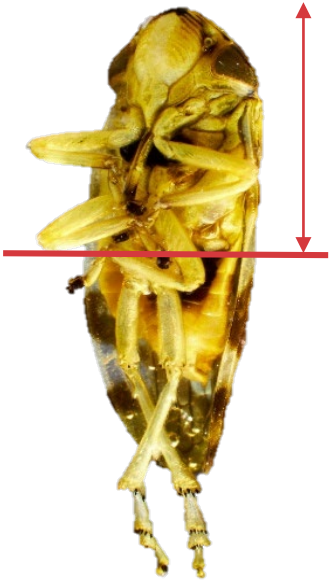


# Nymph and Adult of *N. campestris* on Poaceae (Dar Chichou)





## Detection of *X. fastidiosa* in insects



Extraction of ADN from the upper part of Philaenus and qPCR

Morphological identification by dissection of male genitalia (lower insect part)

Analyze insects, potential vectors to identify whether they are contaminated by the bacteria

No *Xylella* identified in insect until now

- Remember when *Xylella fastidiosa* is detected in the (pre)cibarium of the insect, that does not mean it will be an effective vector for a given plant
- Vector transmissions are necessary.

# Enemies ?

Diptera



Hymenoptera



Not identified yet

## Evaluation of the Risk

- Occurrence of a confirmed vector in Tunisia : *Neophilaenus campestris*
- Occurrence of an important potential vector : *Philaenus tessellatus*, very close in morphology and behavior to *Philaenus spumarius*
- Occurrence of *P. maghresignus* similar to *P. italosignus* a vector too
- Plant trade and exchanges are currently made

What is the necessary measures ?





A photograph of a dense field of yellow daisies with green foliage. In the lower right, a brown beetle is visible. A yellow speech bubble with a green outline is positioned above the beetle, containing the text "Thank you for your interest".

Thank you for  
your interest